



Bioaccumulation Pattern of Heavy Metals in Commercially Important Fishes in and Around Indian Sundarbans

Abhijit Mitra and Rajrupa Ghosh*

Department of Marine Science, University of Calcutta, 35. B.C. Road, Kolkata; Techno India University, India

ARTICLE INFO

Corresponding Author:

Rajrupa Ghosh
rajrupa14@gmail.com

How to cite this article:

Mitra, A., and R. Ghosh. 2014. Bioaccumulation Pattern of Heavy Metals in Commercially Important Fishes in and Around Indian Sundarbans. *Global Journal of Animal Scientific Research*. 2(1): 33-44.

Article History:

Received: 6 February 2014
Received in revised form:
21 February 2014
Accepted: 23 February 2014

ABSTRACT

Concentrations of Zn, Cu, Pb and Cd were determined in edible finfish species (*Polynemus paradiseus*, *Tenuulosa ilisha*, *Liza parsia*, *Liza tade* and *Stolephorus commersonii*) collected from four stations in and around Indian Sundarbans. Levels of the selected heavy metals were determined in the muscle of edible finfish species in the Gangetic delta region using a Perkin-Elmer Sciex ELAN 5000 ICP mass spectrometer and expressed as mg kg⁻¹ dry weight. To determine whether heavy metal concentrations varied significantly between sites and species, Duncan multiple range test was performed. In finfish species the concentrations of Zn, Cu, Pb and Cd ranged from 15.89±0.58 – 124.12 ±1.63 ppm, 13.68±0.83 – 75.91±0.49 ppm, 3.34±0.90–19.89±0.54 ppm and BDL–4.01±0.03 ppm respectively. For Zn and Cu, accumulated metal concentration in Stn. 4 is significantly lower than accumulated metal concentration in Stn. 1 and Stn. 2. For Pb, significant difference between stations was not found. Among the selected species, lowest metal accumulation values was found for *S. commersonii* (p<0.05). The selected heavy metals in finfish muscle (except Zn in *Liza parsia* and *Liza tade* in Stn. 1) were also within the permissible limits for human consumption as indicated by the Food and Agricultural Organization.

Keywords: Finfish; Heavy metals; Indian Sundarbans

©World Science and Research Publications, 2014

INTRODUCTION

The Indian Sundarbans in the lower Gangetic delta, at the apex of Bay of Bengal is recognized as one of the most diversified and productive ecosystems of the Tropics. The deltaic lobe is unique for its wilderness, mangrove gene pool and tiger habitat. However due to intense industrial activities in the upstream zone, and several anthropogenic factors, the western part of the deltaic complex is exposed to pollution from domestic sewage and industrial effluents leading

to serious impacts on biota (Mitra and Choudhury, 1992). The presence of Haldia port-cum-industrial complex in the downstream region of the River Ganga (also known as the Hooghly River) has accelerated the pollution problem with a much greater dimension (Mitra, 1998). The organic and inorganic wastes released from these industries and urban units contain substantial concentrations of heavy metals. The central part of the delta (encompassing the surroundings of Matla River) is relatively less stressful in terms of industrial discharge. Due to siltation of the Bidyadhari channel the area does not receive any water supply from the Hooghly River in the western sector and is therefore tide-fed in nature receiving the tidal flux from the Bay of Bengal (average salinity= ~32 psu). 85 percent of the people in this area consume fishes that are caught from the Gangetic delta region.

The information which is available in the literature suggests that the concentrations of toxic metals in the ecosystem are reaching unprecedented levels. Due to the steady load of contaminated dust in overcrowded cities, the ambient concentrations of toxic metals are now among the highest ever being reported. Metals are elements that accumulate and do not break down further into less harmful constituent (Kennicutt *et al.*, 1992). With increasing urbanization and industrialization, there is a rapid increase of environmental pollution, which is attracting the attention of people around the world. The major sources of contamination in surface water can be traced to industrial discharges, domestic waste disposal and application of agrochemicals on farmlands (Kennicutt *et al.*, 1992). The pollutants like heavy metals after entering into aquatic environment accumulate in tissues and organs of aquatic organisms. The amount of absorption and assembling depends on ecological, physical, chemical and biological condition and the kind of element and physiology of organisms (Jaffa, 1998). These metals ultimate consumed by human, after accumulation by the body of aquatic organisms enter into food chain (Agbozu, 2007). Water - borne chemicals are absorbed in the gills, skin and digestive tract in fish and then transported by the blood to either a storage point such as the bone, or to the liver for transportation. If transported by the liver, it may be stored there, excreted in the bile, or passed back into the blood for possible excretion by the kidney or gills or stored in extra hepatic tissues such as fat (McNicol and Scherer, 1991).

Most of people in West Bengal are fish lover, so, the present paper aims to highlight the concentration of selective heavy metals (Zn, Cu, Pb and Cd) in the muscle tissue of five common finfish species namely *Polynemus paradiseus*, *Tenualosa ilisha*, *Liza parsia*, *Liza tade* and *Stolephorus commersonii* from four stations distributed in two sectors (western and central Indian Sundarbans) of the lower Gangetic region.

Description of the study site

Two sampling sites were selected each in the western and central sectors of Indian Sundarbans, a Gangetic delta at the apex of the Bay of Bengal. The deltaic complex has an area of 9630 sq. Km and houses 102 islands. The western sector of the deltaic lobe receives the snowmelt water of mighty Himalayan glaciers after being regulated through several barrages on the way. The central sector on the other hand, is fully deprived from such supply due to heavy siltation and clogging of the Bidyadhari channel since the late 15th century (Chaudhuri & Choudhury, 1994). The western sector also receives wastes and effluents of complex nature from multifarious industries concentrated mainly in the upstream zone. On this background four sampling stations (two each in western and central sectors) were selected (Table 1 and Fig. 1) to analyze the concentrations of heavy metals in the common edible finfish and shellfish species inhabiting the zone.

Table 1: List of sampling stations with coordinates and salient features

Station	Coordinates	Salient Features
Nayachar Island (Stn.1)	88° 15' 24" E 21° 45' 24" N	It is located in the Hooghly estuary in the western sector of the lower Gangetic delta and faces the Haldia port-cum-industrial complex that houses a variety of industrial units.
Sagar South(Stn.2)	88° 01' 47" E 21° 39' 04" N	Situated at the confluence of the River Hooghly and the Bay of Bengal in the western sector of Indian Sundarbans, the station is an important navigational channel for the major ports of the area.
Gosaba(Stn. 3)	88° 39' 46" E 22° 15' 45" N	Located in the Matla Riverine stretch in the central sector of Indian Sundarbans.
Annpurin Satjelia Island (Stn. 4)	88° 50' 43" E 22° 11' 52" N	Located in the central sector of Indian Sundarbans. Noted for its wilderness and mangrove diversity; selected as our control zone.

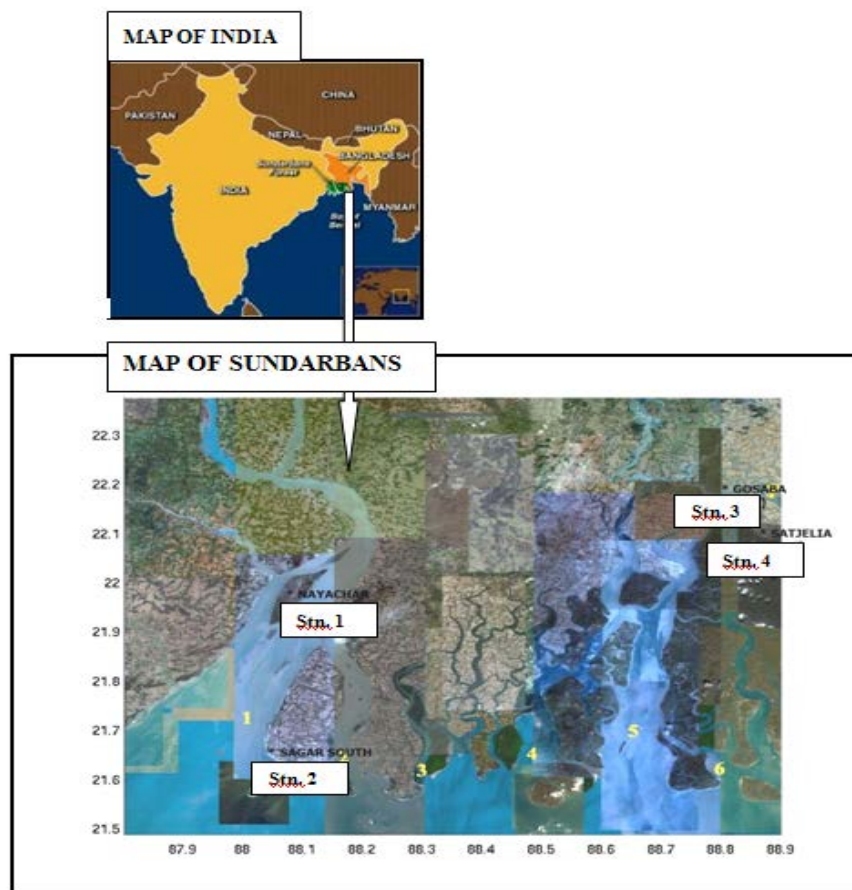


Fig.1. Location of sampling stations

MATERIALS AND METHODS

Sampling of Specimen

Five commonly edible finfish species Paradise fish (*Polynemus paradiseus*), Hilsa (*Tenualosa ilisha*), Parsey (*Liza parsia*), Tade Mullet (*Liza tade*) and Anchovy (*Stoleophorus commersonii*) were collected during low tide condition from the selected stations (Table 1) during a rapid EIA study from 15th October to 30th October, 2013. The collected samples were stored in a stored container, preserved in crushed ice with 1:1 fish to ice ratio and brought to the laboratory for further analysis.

Analysis

Inductively Coupled Plasma – Mass Spectrometry (ICP-MS) is now - a - day accepted as a fast, reliable means of multi-elemental analysis for a wide variety of sample types (Date & Gray, 1988). A Perkin-Elmer Sciex ELAN 5000 ICP mass spectrometer was used for the present analysis. A standard torch for this instrument was used with an outer argon gas flow rate of 15 L/min and an intermediate gas flow of 0.9 L/min. The applied power was 1.0 kW. The ion settings were standard settings recommended, when a conventional nebulizer/spray is used with a liquid sample uptake rate of 1.0 mL/min. A Moulinex Super Crousty microwave oven of 2450 MHz frequency magnetron and 1100 Watt maximum power Polytetrafluoroethylene (PTFE) reactor of 115 ml volume, 1 cm wall thickness with hermetic screw caps, were used for the digestion of the collected biological samples. All reagents used were of high purity available and of analytical reagent grade. High purity water was obtained with a Barnstead Nanopure II water-purification system. All glasswares were soaked in 10% (v/v) nitric acid for 24 h and washed with deionised water prior to use.

The analyses were carried out on composite samples of 10 specimens of each species having uniform size. This is a measure to reduce possible variations in metal concentrations due to size and age. 20 mg composite sample from each species of finfish and shellfish were weighed and successively treated with 4 ml aqua regia, 1.5 mL HF and 3 ml H₂O₂ in a hermetically sealed PIFE reactor, inside a microwave oven, at power levels between 330-550 Watt, for 12 min to obtain a clear solution. The use of microwave-assisted digestion appears to be very relevant for sample dissolution, especially because it is very fast (Nadkarni, 1984; Matusiewicz & sturgeon, 1989; De la Guardia, 1990). After digestion, 4 ml H₂BO₃ was added and kept in a hot water bath for 10 min, diluted with distilled water to make up the volume to 50 ml. Taking distilled water in place of biological samples and following all the treatment steps described above the blank process was prepared. The final volume was made up to 50 ml. Finally, the samples and process blank solutions were analyzed by ICP-MS. All analyses were done in triplicate and the results were expressed with standard deviation.

The accuracy and precision of our results were checked by analyzing standard reference material (SRM, Dorm-2). The results indicated good agreement between the certified and the analytical values (Table 2 and 3).

Statistical analysis

A logarithmic transformation was done on the data to improve normality. Duncan multiple range tests was performed to assess whether heavy metal concentrations varied significantly between sites and species; possibilities less than 5% ($p < 0.05$) were considered statistically

significant. All statistical calculations were performed with SPSS 21.0 for Windows. Superscripts were used to show the statistically significant difference between stations and species.

RESULTS AND DISCUSSION

The species-wise variation was not uniform for all the metals. Zn accumulated as per the order *Liza parsia* > *Liza tade* > *Tenualosa ilisha* > *Polynemus paradiseus* > *Stolephorus commersonii* (Table 2a, 2b and Fig. 2). Cu accumulated as per the order *Liza parsia* > *Tenualosa ilisha* > *Polynemus paradiseus* > *Liza tade* > *Stolephorus commersonii* (Table 2a, 2b and Fig. 3). Pb accumulated as per the order *Liza parsia* > *Liza tade* > *Tenualosa ilisha* > *Polynemus paradiseus* > *Stolephorus commersonii* (Table 3a, 3b and Fig. 4). Cd was BDL in all the stations except station 1, where the order was *Liza parsia* > *Liza tade* > *Tenualosa ilisha* (Table 3a, 3b and Fig. 5). For Zn and Cu, an accumulated metal concentration in Stn. 4 is significantly lower than accumulated metal concentrations in Stn 1 and Stn 2. For Pb, significantly station difference between stations was not found. Between all studies fish species, lowest metal accumulation values was found for *S. commersonii* ($p < 0.05$).

Heavy metals are non-biodegradable and once discharged into water bodies; they can be absorbed on sediments particles or accumulated in aquatic organisms (Kotze *et al.*, 1999). Especially fish, which in turn may enter into the human metabolism through consumption causing serious health hazards (Jeziarska and Witeska, 2001). Accumulation of metal in different species may be the function of their respective. Fish species mostly absorbed heavy metals from its feeding diets, sediments and surrounding waters resulting to their accumulation in reasonable amounts (McCarthy and Shugart, 1990). Microhabitat utilization, feeding habits, age, sex and fish species also determine the accumulation pattern of heavy metals (Kotze *et al.*, 1999). Bioaccumulation is species-dependent and therefore feeding habits and life style can be strongly related to the sediment exposure (Chen and Chen, 1999). On the other hand, bioavailability of metals can be influenced by inorganic and organic factors that control metal speciation and thereby bioaccumulation (Henry *et al.*, 2004). The metal accumulation in different fish organs depends on their physiological role, behavior and feeding habits, as well as regulatory ability, as reported by Clearwater *et al.* (2002). Other factors, such as sex and size may also influence metal bioaccumulation (Al-Yousuf *et al.*, 1999; Canli and Atli, 2003).

Heavy metals are stable and persistent environmental contaminants of aquatic environments. They occur in the environment both as a result of natural processes and as pollutants from human activities (Garcia-Montelongo *et al.*, 1994; Jordao *et al.*, 2002). Some metals like Zn and Cu, which are required for metabolic activity in organisms, lie in the narrow "window" between their essentiality and toxicity. Other heavy metals like Cd and Pb, may exhibit extreme toxicity even at low levels under certain conditions, thus necessitating regular monitoring of sensitive aquatic environments (Cohen *et al.*, 2001; Fergusson, 1990; Peerzada *et al.*, 1990).

From an environmental point of view, coastal zones can be considered as the geographic space of interaction between terrestrial and marine ecosystems that is of great importance for the survival of a large variety of plants, animals and marine species (Castro *et al.*, 1999). The coastal zone receives a large amount of metal pollution from agricultural and industrial activity (Usero *et al.*, 2005). Adverse anthropogenic effects on the coastal environment include eutrophication, heavy metals, organic and microbial pollution and oil spills (Boudouresque & Verlaque, 2002).

Table 2a. Zn and Cu concentrations (in ppm dry wt.) in finfish muscles (station wise)*

Species	Zn				Cu			
	Stn. 1	Stn. 2	Stn. 3	Stn. 4	Stn. 1	Stn. 2	Stn. 3	Stn. 4
<i>Polynemus paradiseus</i>	76.89 ^d ± 1.20	67.89 ^c ± 1.56	41.26 ^d ± 1.49	15.89 ^d ± 0.58	62.73 ^c ± 0.45	44.12 ^d ± 1.15	21.81 ^d ± 0.19	15.34 ^d ± 0.91
<i>Tenualosa ilisha</i>	93.46 ^c ± 1.64	78.12 ^b ± 1.34	46.35 ^c ± 1.34	18.22 ^b ± 0.35	66.12 ^b ± 0.48	46.56 ^c ± 1.48	25.57 ^c ± 0.38	17.98 ^c ± 0.33
<i>Liza parsia</i>	124.12 ^a ± 1.63	94.63 ^a ± 1.21	61.21 ^a ± 1.67	27.67 ^a ± 1.89	75.91 ^a ± 0.49	54.39 ^a ± 1.24	28.12 ^a ± 0.67	21.01 ^a ± 0.73
<i>Liza tade</i>	103.45 ^b ± 1.49	91.25 ^a ± 1.45	53.98 ^b ± 1.23	25.45 ^a ± 1.12	43.89 ^d ± 0.36	50.38 ^b ± 1.67	27.91 ^b ± 0.55	18.65 ^b ± 0.88
<i>Stolephorus commersonii</i>	45.03 ^c ± 1.02	25.16 ^d ± 1.63	19.26 ^e ± 1.15	16.32 ^c ± 0.65	16.12 ^c ± 0.35	14.90 ^e ± 0.73	13.68 ^e ± 0.83	13.91 ^e ± 0.91
WHO (1989) level for Zn and Cu in food		100 ppm					30 ppm	
FAO (1992) level for Zn and Cu in fish		30-100 ppm					10-100 ppm	

*Means in a whole column with different superscripts (a-e) are significantly different ($p < 0.05$, Duncan multiple range test).

Table 2b. Zn and Cu concentrations (in ppm dry wt.) in finfish muscles (species wise)*

Species	Zn				Cu			
	Stn. 1	Stn. 2	Stn. 3	Stn. 4	Stn. 1	Stn. 2	Stn. 3	Stn. 4
<i>Polynemus paradiseus</i>	76.89 ^a ± 1.20	67.89 ^b ± 1.56	41.26 ^c ± 1.49	15.89 ^d ± 0.58	62.73 ^a ± 0.45	44.12 ^b ± 1.15	21.81 ^c ± 0.19	15.34 ^d ± 0.91
<i>Tenualosa ilisha</i>	93.46 ^a ± 1.64	78.12 ^b ± 1.34	46.35 ^c ± 1.34	18.22 ^d ± 0.35	66.12 ^a ± 0.48	46.56 ^b ± 1.48	25.57 ^c ± 0.38	17.98 ^d ± 0.33
<i>Liza parsia</i>	124.12 ^a ± 1.63	94.63 ^b ± 1.21	61.21 ^c ± 1.67	27.67 ^d ± 1.89	75.91 ^a ± 0.49	54.39 ^b ± 1.24	28.12 ^c ± 0.67	21.01 ^d ± 0.73
<i>Liza tade</i>	103.45 ^a ± 1.49	91.25 ^b ± 1.45	53.98 ^c ± 1.23	25.45 ^d ± 1.12	43.89 ^b ± 0.36	50.38 ^a ± 1.67	27.91 ^c ± 0.55	18.65 ^d ± 0.88
<i>Stolephorus commersonii</i>	45.03 ^a ± 1.02	25.16 ^b ± 1.63	19.26 ^c ± 1.15	16.32 ^d ± 0.65	16.12 ^a ± 0.35	14.90 ^b ± 0.73	13.68 ^d ± 0.83	13.91 ^c ± 0.91
WHO (1989) level for Zn and Cu in food		100 ppm				30 ppm		
FAO (1992) level for Zn and Cu in fish		30-100 ppm				10-100 ppm		

*Means in a whole column with different superscripts (a-e) are significantly different ($p < 0.05$, Duncan multiple range test).

The discharge of these wastes without adequate treatment often contaminate the estuarine and coastal water with conservative pollutants (like heavy metals), many of which accumulate in the tissues of resident organisms like fishes, oysters, crabs, shrimps, seaweeds *etc.* In many parts of the world, especially in coastal areas and on smaller islands, fish is a major part of food, which supplies all essential elements required for life processes in a balanced manner (Iyengar, 1991). Hence, it is important to investigate the levels of heavy metals in these organisms to assess whether the concentration is within the permissible level and will not pose any hazard to the consumers (Krishnamurti & Nair 1999).

Table 3a. Pb and Cd concentrations (in ppm dry wt.) in finfish muscles (station wise) *

Species	Pb				Cd			
	Stn. 1	Stn. 2	Stn. 3	Stn. 4	Stn. 1	Stn. 2	Stn. 3	Stn. 4
<i>Polynemus paradiseus</i>	10.99 ^c ± 0.26	9.71 ^c ± 0.35	9.04 ^d ± 0.70	7.20 ^d ± 0.48	BDL	BDL	BDL	BDL
<i>Tenualosa ilisha</i>	14.01 ^b ± 0.15	13.12 ^b ± 0.98	11.30 ^c ± 0.65	9.39 ^c ± 0.53	1.35 ^c ± 0.82	BDL	BDL	BDL
<i>Liza parsia</i>	19.89 ^a ± 0.54	15.16 ^a ± 0.53	14.85 ^b ± 0.67	13.92 ^a ± 0.46	4.01 ^a ± 0.03	BDL	BDL	BDL
<i>Liza tade</i>	15.77 ^b ± 0.45	13.51 ^b ± 0.47	15.79 ^a ± 0.90	11.65 ^b ± 0.42	1.95 ^b ± 0.16	BDL	BDL	BDL
<i>Stolephorus commersonii</i>	4.65 ^d ± 0.04	3.69 ^d ± 0.89	5.01 ^e ± 0.44	3.34 ^e ± 0.90	BDL	BDL	BDL	BDL
WHO (1989) level for Pb and Cd in food		2 ppm					1 ppm	
FAO (1992) level for Pb and Cd in fish		0.5 – 6.0 ppm					0.05 – 5.5 ppm	

BDL means below detectable level

*Means in a whole column with different superscripts (a–e) are significantly different ($p < 0.05$, Duncan multiple range test).**Table 3b. Pb and Cd concentrations (in ppm dry wt.) in finfish muscles (species wise)***

Species	Pb				Cd			
	Stn. 1	Stn. 2	Stn. 3	Stn. 4	Stn. 1	Stn. 2	Stn. 3	Stn. 4
<i>Polynemus paradiseus</i>	10.99 ^a ± 0.26	9.71 ^b ± 0.35	9.04 ^b ± 0.70	7.20 ^c ± 0.48	BDL	BDL	BDL	BDL
<i>Tenualosa ilisha</i>	14.01 ^a ± 0.15	13.12 ^b ± 0.98	11.30 ^c ± 0.65	9.39 ^d ± 0.53	1.35± 0.82	BDL	BDL	BDL
<i>Liza parsia</i>	19.89 ^a ± 0.54	15.16 ^b ± 0.53	14.85 ^c ± 0.67	13.92 ^d ± 0.46	4.01 ± 0.03	BDL	BDL	BDL
<i>Liza tade</i>	15.77 ^a ± 0.45	13.51 ^b ± 0.47	15.79 ^a ± 0.90	11.65 ^c ± 0.42	1.95 ± 0.16	BDL	BDL	BDL
<i>Stolephorus commersonii</i>	4.65 ^b ± 0.04	3.69 ^{cd} ± 0.89	5.01 ^a ± 0.44	3.34 ^d ± 0.90	BDL	BDL	BDL	BDL
WHO (1989) level for Pb and Cd in food		2 ppm					1 ppm	
FAO (1992) level for Pb and Cd in fish		0.5 – 6.0 ppm					0.05 – 5.5 ppm	

BDL means below detectable level

*Means in a whole column with different superscripts (a–e) are significantly different ($p < 0.05$, Duncan multiple range test).

Heavy metals have contaminated the aquatic environment in the present century due to intense industrialization and urbanization. The Gangetic delta is no exception to this usual trend. The rapid industrialization and urbanization of the city of Kolkata (formerly known as Calcutta), Howrah and the newly emerging Haldia complex in the maritime state of West Bengal has caused considerable ecological imbalance in the adjacent coastal zone (Mitra & Choudhury, 1992; Mitra,

1998). The Hooghly estuary, situated on the western sector of the Gangetic delta receives drainage from these adjacent cities, which have sewage outlets into the estuarine system. The chain of factories and industries situated on the western bank of the Hooghly estuary is a major cause behind the gradual transformation of this beautiful ecotone into stinking cesspools of the megapolis (Mitra & Choudhury, 1992). The lower part of the estuary has multifarious industries such as paper, textiles, chemicals, pharmaceuticals, plastic, shellac, food, leather, jute, tyres and cycle rims (UNEP, 1982). These units are point sources of heavy metals in the estuarine water. Due to toxic nature of certain heavy metals, these chemical constituents interfere with the ecology of a particular environment and on entering into the food chain they cause potential health hazards, mainly to human beings. It was reported by several workers that the discharge of heavy metals into the sea through rivers and streams results in the accumulation of pollutants in the marine environment especially within fishes (Yusof *et al.*, 1994). Thus fishes can be used for monitoring potential risk to humans because these are directly consumed by a large population (Subramanian & Sukumar, 1988).

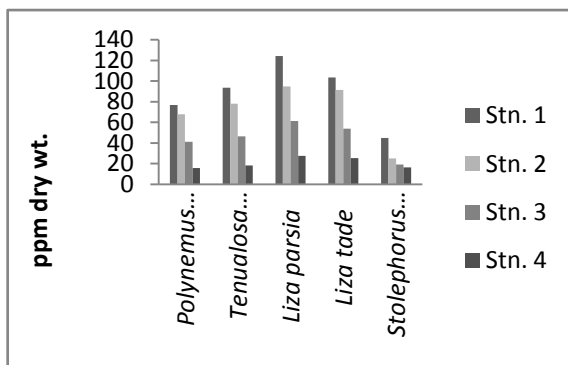


Fig. 2. Zn concentrations in finfish muscle

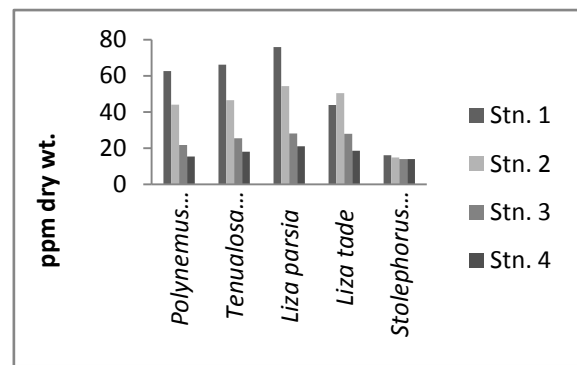


Fig. 3. Cu concentrations in finfish muscle

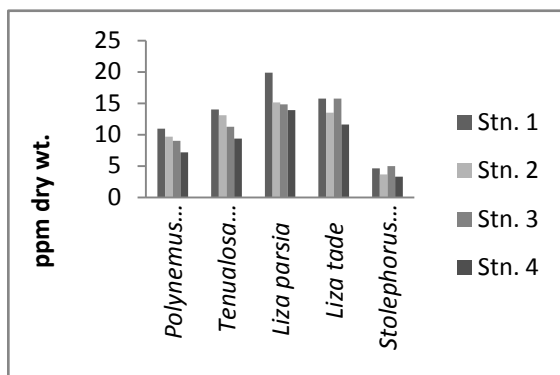


Fig.4. Pb concentrations in finfish muscle

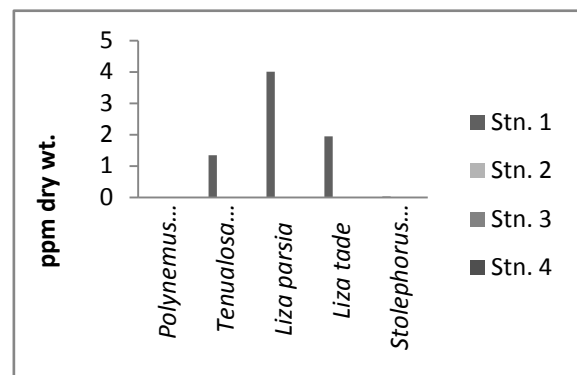


Fig. 5. Cd concentrations in finfish muscle

The selected finfish species in the present study have different food preference and different behavioral pattern e.g., *Liza parsia*, *Liza tade*, *Polynemus paradiseus*, and *Stolephorus commersonii* are resident fish species in the study area, while *Tenualosa ilisha* exhibit migration

from coastal region (~ salinity = 20 psu) to freshwater system in the upstream zone of the River Ganga for breeding. These factors may be attributed to species-wise variation of heavy metals in the study zone. The spatial variation of bioaccumulation followed the order station 1 > station 2 > station 3 > station 4, which may be related to different degree of contamination in different location. All the heavy metals (except Zn in *Liza parsia* in station 1) were found to be lower than the recommended maximum level allowed in food as prescribed by the World Health Organization (WHO, 1989). Furthermore the selected heavy metals in finfish muscle (except Zn in *Liza parsia* in station 1) were also within the permissible limits for human consumption as indicated by the Food and Agricultural Organization (FAO, 1992).

Of the four metals studied in the present work, Zn and Cu are essential elements while Pb and Cd are non-essential elements for most of the living organisms. The concentrations of Zn and Cu in all the finfish and shellfish species were relatively higher, compared to the concentration of other metals in same samples. It can be explained because these metals (Cu and Zn) are essential elements required by animals for metabolic process. Zn and Cu appear to diffuse passively (probably as a soluble complex) the gradients created by adsorption of membrane surfaces and are found in blood proteins metallothioneins. Carbonell and Tarazona (1994) concluded that different tissues of aquatic animals provide and/or synthesize nonexchangeable binding sites resulting in different accumulation levels. The primary sources of Zn in the present geographical locale are the galvanization units, paint manufacturing units and pharmaceutical processes, which are mainly concentrated in the Haldia industrial sector (opposite to station 1). Reports of high concentrations of Zn were also highlighted in the same environment by earlier workers (Mitra and Choudhury, 1992; Mitra and Choudhury, 1993; Mitra, 1998).

The main sources of Cu in the coastal waters are antifouling paints (Goldberg, 1975), particular type of algaecides used in different aquaculture farms, paint manufacturing units, pipe line corrosion and oil sludges (32 to 120 ppm). Ship bottom paint has been found to produce very high concentration of Cu in sea water and sediment in harbours of Great Britain and southern California (Bellinger & Benham, 1978; Young *et al.*, 1979). In the present study area, the major source of Cu is the antifouling paints used for conditioning fishing vessels and trawlers apart from industrial discharges (that is predominant around station 1). This is the reason why Cu was detected in the fish samples of stations 3 and 4, even there are no existence of industries. The complete siltation of the Bidyadhari River also does not permit the industrial effluents released in the Hooghly River to mix with the rivers in the central sector of the deltaic complex (location zone of stations 3 and 4).

Pb is a toxic heavy metal, which finds its way in coastal waters through the discharge of industrial waste waters, such as from painting, dyeing, battery manufacturing units and oil refineries (Mitra, 1998). Antifouling paints used to prevent growth of marine organisms at the bottom of the boats and trawlers also contain Pb as an important component. These paints are designed to constantly leach toxic metals into the water to kill organisms that may attach to bottom of the boats, which ultimately is transported to the sediment and aquatic compartments. Lead also enters the oceans and coastal waters both from terrestrial sources and atmosphere and the atmospheric input of lead aerosols can be substantial. Station 1 is exposed to all these activities being proximal to the highly urbanized city of Kolkata, Howrah and the newly emerging Haldia port - cum - industrial complex, which may be attributed to high Pb concentrations in the finfish species.

The main sources of Cd in the present geographical locale are electroplating, manufacturing of Cd alloys, production of Ni-Cd batteries and welding (Mitra, 1998). No trace of Cd was recorded

in the fish muscle from stations 3 and 4, which are located almost in industry-free zone surrounded by mangrove vegetation.

A part from industrial discharges, significant difference in metal concentrations in biological samples between stations ($p < 0.01$) may also be related to contrasting physico-chemical characteristics between the western and central part of the Gangetic delta. The western part of the Gangetic delta is connected to Himalayan glacier through Bhagirathi River. Researchers pointed out that the glaciers in the Himalayan range are melting at the rate of 23 m/yr (Hasnain, 1999; 2000; 2002). This along with Farakka discharge has resulted in gradual freshening of the system (Mitra et al., 2009), which has role in elevation of dissolved metal level in the aquatic phase by way of lowering the salinity, pH and enhancing the process of dissolution of metallic species (Mitra, 1998). The central sector on contrary is derived from freshwater supply of Ganga-Bhagirathi system, and the Matla River is now tide fed with an increasing trend of salinity. This results in the precipitation of dissolved metals on the sediment bed (an event of compartmentation due to variation in metallic species) making the availability of metal low to the tissue of fish species thriving in the system (Mitra and Choudhury, 1992; Mitra and Choudhury, 1993; Mitra, 1998). Similar result has been found by many researchers (Ambedkar and Muniyan, 2011; Kamaruzzaman et al., 2011; Heidary et al., 2011)

This study reflects in some zones in and around Indian Sundarbans ecosystem, the bioaccumulation of certain heavy metal has touched the alarming level. Hence, strict regulations should be implemented to control industrial waste from the point sources both in the upstream and downstream areas of the Indian Sundarban deltaic complex.

REFERENCE

- Ambedkar, G., and M. Muniyan .1999. Accumulation of metals in the five commercially important freshwater fishes available in vellar river, Tamil Nadu, India. *Scholars Research Library*. 3(3):261-264.
- Al-Yousuf, M.H., M.S. El-Shahawi, and S.M. Al-Ghais. 1999. Trace metals in liver, skin and muscle of *Lethrinus lentjan* fish species in relation to body length and sex. *Sci. Total Environ.* 256:87-94. DOI:10.1016/S0048-9697(99)00363-0.
- Bellinger, E., B. Benhem .1978. The levels of metals in Dockyard sediments with particular reference to the contributions from ship bottom paints. *Environ. Poll. Assess.* 15(1): 71-81.
- Boudouresque, C.F., M. Verlaque. 2002. Biological pollution in the Mediterranean Sea: invasive versus introduced macrophytes. *Mar. Poll. Bull.* 44: 32-38.
- Canli, M. and G. Atli .2003. The relationships between heavy metal (Cd, Cr, Cu, Fe, Pb, Zn) levels and the size of six Mediterranean fish species. *Environ. Pollut.* 121: 129-136. DOI:10.1016/S0269-7491(02)00194-X.
- Carbonell, G., and J.V. Tarazona .1994. Toxicokinetics of Cu. *Aqua. Toxicol.* 29: 213-221.
- Castro, H., P.A. Aguilera, J.L. Martinez, and E.L. Carrique. 1999. Differentiation of clams from fishing areas an approximation to coastal quality assessment. *Environ. Monit. Assess.* 54: 229-237.
- Chaudhuri, A.B., and A. Choudhury. 1994. Mangroves of the Sundarbans, India, Vol-I., first ed. IUCN.
- Chen, M.H., and C.Y. Chen.1999. Bioaccumulation of sediment-bound heavy metals in grey mullet, *Liza macrolepis*. *Mar. Pollut. Bull.* 39:239-244. DOI:10.1016/S0025-326X(99)00027-2.
- Clearwater, S.J., A.M. Farag, and J.S. Meyer. 2002. Bioavailability and toxicity of dietborne copper and zinc to fish. Comparative biochemistry and physiology part C: *Toxicol. Pharmacol.* 132:269-313. DOI:10.1016/S1532-0456(02)00078-9.
- Cohen, T., S. Hee, and R. Ambrose. 2001. Trace metals in fish and invertebrates of three

- California Coastal Wetlands. *Mar. Poll. Bull.* 42: 232-242.
- Date, A.R., and A.L. Gray (Eds.) .1988. Applications of Inductively Coupled Plasma Source Mass Spectrometry, Blackie, Glasgow.
- De la Guardia, M. (Ed.) .1990. Empleo de los Hornos de Microondas en Química, University of Valencia, Spain.
- Enoyer, E.R., .1992. Semi-quantitative analysis of environmental materials by laser-sampling inductively coupled plasma mass spectrometry. *J. Anal. Atom. Spectro.* 7: 1187.
- Fergusson, J.E. 1990. The heavy elements: Chemistry, environmental impact and health effects, Pergamon Press, New York. p: 614.
- FAO .1992. FAO/WHO, Food standard programme. 2nd ed. Codex Alimentarius Commission. Vol. 1. pp: 114-190.
- Garcia-Montelongo, F., L.Galindo, M.S. Larrechi, X. Rius .1994. Heavy metals in three fish species from the coastal waters of Santa Cruz de Tenerife (Canary Islands). *Scientia Marina.* 58: 179-183.
- Goldberg, E.D. 1975. The mussel watch – a first step in global marine monitoring. *Mar. Poll. Bull.* 6: 111.
- Grosell, M., K.V. Brix. 2005. Introduction to the special issue on mechanisms in metal toxicology. *Aqua. Toxicol.* 72: 3-4.
- Hasnain, S.I.,.1999. Himalayan Glaciers: Hydrology and Hydrochemistry. First ed, Allied Publication Limited, New Delhi.
- Hasnain, S.I. 2000. Status of the Glacier Research in the HKH region. ICIMOD, Kathmandu, Nepal.
- Hasnain, S.I. 2002. Himalayan Glaciers Meltdown: Impact on South Asian Rivers. IAHS; 7: 274.
- Heidary, S., J. Imanpour Namin, and F. Monsefrad. 2011. Bioaccumulation of heavy metals Cu, Zn, and Hg in muscles and liver of the stellate sturgeon (*Acipenser stellatus*) in the Caspian Sea and their correlation with growth parameters. *Iranian Journal of Fisheries Sciences.* 11(2): 325-337.
- Henry, F., R. Amara, L. Courcot, D. Lacouture, and M.L. Bertho. 2004. Heavy metals in four fish species from the French coast of the Eastern English Channel and Southern bight of the North Sea. *Environ. Int.* 30:675-683. DOI: 10.1016/j.envint.2003.12.007.
- Islam, M.D., M. Tanaka. 2004. Impact of pollution on coastal and marine ecosystems including coastal and marine fisheries and approach for management: a review and synthesis. *Mar. Poll. Bull.* 48: 624-649.
- Iyengar, G.V. 1991. Milestones in biological trace elements research. *Sci. Total Environ.* 1: 100.
- Jaffa, M., M. Ashraf, and Rasool. 1998. Heavy metals contents insome selected local freshwater fish and relevant water. *Pakistan Journal of Scientific and Industrial Research.* 31:189-193.
- Jezierska, B., and M. Witeska .2001. The metal uptake and accumulation in fish living in polluted waters. Department animal Physiology. University of podlasie. Pursa. 12. 108-110 Siedlce. Polant. pp :107.
- Jordao, C. P., M.G. Pereira, C. R. Bellato, J.L. Pereira, A.T. Matos. 2002. Assessment of water systems for contaminants from domestic and industrial sewages. *Environ. Monit. Assess.* 79: 75-100.
- Kamaruzzaman, B.Y, M.C. Ong, and S.Z. Rina. 2011. Heavy metal accumulation in commercially important fishes of South West Malaysian Coast. *Research Journal of Environmental Science.* 5(6): 595-602.
- Kennicutt, M.C, T.L. Wade, and B.J. Presley .1992. Texas A&M University. Assessment of Sediment Contamination in Casco Bay. Casco Bay Estuary Project. p:113.
- Kotze, P., H.H. Du Preez, and J.H. Van Vuren. 1999. Bioaccumulation of copper and zinc in *Oreochromis mossambicus* and *Clarias gariepinus*, from the Olifants River, Mpumalanga. *South Africa. Water SA.* 25 (1) 99-110.
- Krishnamurti J.A. and R.V. Nair. 1999. Concentration of metals in shrimps and crabs from Thane- Bassein creek system, Maharashtra. *Ind. J. Mar. Sci.* 28: 92-95.
- Matusiewicz, H.,and R.E. Sturgeon.1989. Present status of microwave sample dissolution and decomposition for elemental analysis. *Prog. Anal. Spectro.* 12: 21.
- McCarthy, J.F., and L.R. Shugart. 1990. Biomarkers of environmental Contamination Lewis Publishers, Chelsea, Mich. 3-16.
- McNicol, R.E., and E. Scherer.1991. Behavioral responses of lake whitefish (*Coregonus clupeaformis*) to cadmium during preference-avoidance testing. *Environ. Toxicol. Chem.* 10: 225-234.
- Mitra, A., A. Choudhury. 1992. Trace metals in macrobenthic molluscs of the Hooghly estuary. *India. Mar. Poll. Bull. UK.* 26 (9): 521-522.
- Mitra, A., and A. Choudhury. 1993. Seasonal variations in metal content in the gastropod

Nerita articulata (Gould). *Ind. J. of Env.Hlth. NEERI*. 35 (1): 31-35.

- Mitra, A. 1998. Status of coastal pollution in West Bengal with special reference to heavy metals. *J. Ind. Ocn. Stud.* 5(2): 135 -138.
- Mitra, A., K. Banerjee, K. Sengupta, and A. Gangopadhyay (2009) Pulse of Climate change in Indian Sundarbans: A myth or reality? *Natl. Acad. Sci. Lett.* 32(2):1-2.
- Nadkarni, R.A. .1984. Applications of microwave oven sample dissolution in analysis. *Anal. Chem.* 56: 22-33.
- Peerzada, N., L. McMorro, S. Skiliros, M. Guinea, and P. Ryan. 1990. Distribution of heavy metals in Gove Harbour, Northern Territory, Australia. *Sci.Total Environ.* 92: 1-12.
- Romeo, M., Y. Siaub, Z. Sidoumou, M. Gnassia-Barelli. 1999. Heavy metal distribution in different fish species from the Mauritania coast. *Sci. Total Environ.* 232: 169-175.
- Subramanian R, and A. Sukumar. 1988. Biological reference materials and analysis of toxic elements. *Fresenius Z Anal. Chem.* 332: 623-626.
- UNEP. 1982. Pollution and the marine environment in the Indian Ocean. UNEP Regional Seas Programme Activity Centre. Geneva, Switzerland.
- Usero, J., J. Morillo, and I. Graccia. 2005. Heavy metal concentrations in molluscs from the Atlantic coast of southern Spain. *Chemosphere.* 59: 1175-1181.
- World Health Organization. 1989. Heavy metals environmental aspects. Environmental Health Criteria. No. 85. Geneva, Switzerland.
- Young, D.R., G.V. Alexander, and D. McDermott-Ehrlich. 1979. Vessel related contamination of southern California harbours by copper and other metals. *Marine Pollution Bulletin.* 10: 50-56.
- Yusof AM, N.F. Yanta, and A.K.H. Wood. 2004. The use of bivalves as bio-indicators in the assessment of marine pollution along a coastal area. *J. Radioanal. Nuclear Chem.* 259(1): 119-127.